



ELSEVIER

Perspective

Dynamic functional connectivity in the static connectome of *Caenorhabditis elegans*Steven W. Flavell¹ and Andrew Gordus²

Abstract

A hallmark of adaptive behavior is the ability to flexibly respond to sensory cues. To understand how neural circuits implement this flexibility, it is critical to resolve how a static anatomical connectome can be modulated such that functional connectivity in the network can be dynamically regulated. Here, we review recent work in the roundworm *Caenorhabditis elegans* on this topic. EM studies have mapped anatomical connectomes of many *C. elegans* animals, highlighting the level of stereotypy in the anatomical network. Brain-wide calcium imaging and studies of specified neural circuits have uncovered striking flexibility in the functional coupling of neurons. The coupling between neurons is controlled by neuromodulators that act over long timescales. This gives rise to persistent behavioral states that animals switch between, allowing them to generate adaptive behavioral responses across environmental conditions. Thus, the dynamic coupling of neurons enables multiple behavioral states to be encoded in a physically stereotyped connectome.

Addresses

¹ Picower Institute for Learning and Memory, Department of Brain & Cognitive Sciences, Massachusetts Institute of Technology, Cambridge, MA, USA

² Department of Biology, Solomon H. Snyder Department of Neuroscience, Johns Hopkins University, Baltimore, MD, USA

Corresponding authors: Flavell, Steven W (flavell@mit.edu); Gordus, Andrew (agordus@jhu.edu)

Current Opinion in Neurobiology 2022, 73:102515

This review comes from a themed issue on **Neurobiology of behavior**

Edited by **Tiago Branco** and **Mala Murthy**

For a complete overview see the [Issue](#) and the [Editorial](#)

Available online xxx

<https://doi.org/10.1016/j.conb.2021.12.002>

0959-4388/© 2021 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Introduction

As animals navigate their environments, they are bombarded by a constant stream of diverse sensory cues. It is therefore essential that their nervous systems extract behaviorally meaningful information from the

environment and generate adaptive responses to these stimuli. Understanding how sensorimotor circuits are able to flexibly process sensory cues, depending on context and experience, has been the subject of intense investigation in recent decades. A key issue in understanding these circuits will be to resolve how neuronal activity in a defined anatomical connectome can be modulated over short and long timescales to allow for dynamic sensorimotor integration.

Studies of mammalian internal states and cognitive control have revealed the broad organization of neural circuits that underlie flexible sensorimotor processing. Behavioral responses to sensory cues are profoundly influenced by an animal's internal state — for example, whether it is awake, asleep, hungry, or thirsty. The ascending neuromodulatory systems, including the noradrenergic and cholinergic systems, play critical roles in generating these states. Correspondingly, many of these neuromodulators have been shown to impact sensory responses in higher brain regions that receive modulatory afferents [1–3]. Sensory processing is also flexible over faster timescales, for example in the context of top-down (or “executive”) control over decision-making. Studies of these higher cognitive functions have shown that neurons in specific cortical areas, most prominently areas in prefrontal cortex, often represent task-relevant variables, rather than faithfully encoding sensory stimuli, like neurons in primary sensory cortical areas [4,5]. These representations can rapidly change depending on the demands of the task and provide a way to link behaviorally relevant inputs to sensorimotor transformations elsewhere. These studies continue to provide important insights into the flexible neural dynamics that underlie sensory processing, but due to the complexity of mammalian circuits, they have not yet established clear links between the physical connectivity of neural circuits and flexible changes in neural circuit function and behavior.

In recent years, the nematode *Caenorhabditis elegans* has emerged as a popular system for the investigation of flexible sensory processing and behavior. Studies of *C. elegans* benefit from its experimental tractability: its nervous system contains exactly 302^a neurons

^a While critical for development, the CANL/R neurons lack synapses with any neurons.

connected through a fully-defined connectome [6]. Moreover, a robust genetic toolset enables neural circuit analysis with single-cell precision in this simple nervous system. While early studies of *C. elegans* behavior focused on the neural circuits underlying hard-wired, innate behaviors, subsequent work revealed a surprising degree of flexibility in how this animal responds to sensory cues in its environment. Modern systems neuroscience tools applied in this simple system have now begun to reveal the basis of this flexibility. Here, we review this recent progress and provide an outlook on what remains to be discovered in this small, flexible nervous system.

Dynamic neuronal activity occurs in a largely invariant physical network

While the invariance of neuron identity in *C. elegans* is well-established, evidence that this extends to synaptic stereotypy has only recently been addressed. Of the 302^a neurons identified in the hermaphrodite, 294 are shared between hermaphrodites and males, with males having an additional 91 neurons and 39 muscles, largely devoted to male mating behaviors [7]. Circuits devoted to shared behaviors, like locomotion and chemotaxis, are largely conserved, although there are differences in how male-specific neurons synapse onto these shared networks.

The architecture of the *C. elegans* network has been carefully analyzed. The distributions of the neurons' degrees of connectivity in the electrical and chemical synaptic networks are fairly scale-free, with tails that follow power-law distributions [8]. This means that both networks have “small world” properties like other scale-free networks, such as social networks and the worldwide web, where a small number of nodes serve as “hubs” for the majority of information traffic. The neurons themselves can be roughly divided into three categories: sensory neurons, interneurons, and motor neurons. On average, the minimum number of chemical synapses crossed from sensory to motor neurons is roughly 3. This means that, on average, the shortest path from sensory neuron to motor neuron will pass through two interneurons. Within the interneuron class are a small number of hub interneurons called the “command neurons” that possess the most synapses in the network, and that form their own, highly interconnected network that controls forward and reverse movement [9–11]. These neurons are pre-synaptic to body motor neurons, and post-synaptic to a large class of interneurons that are themselves post-synaptic to sensory neurons (Figure 1a).

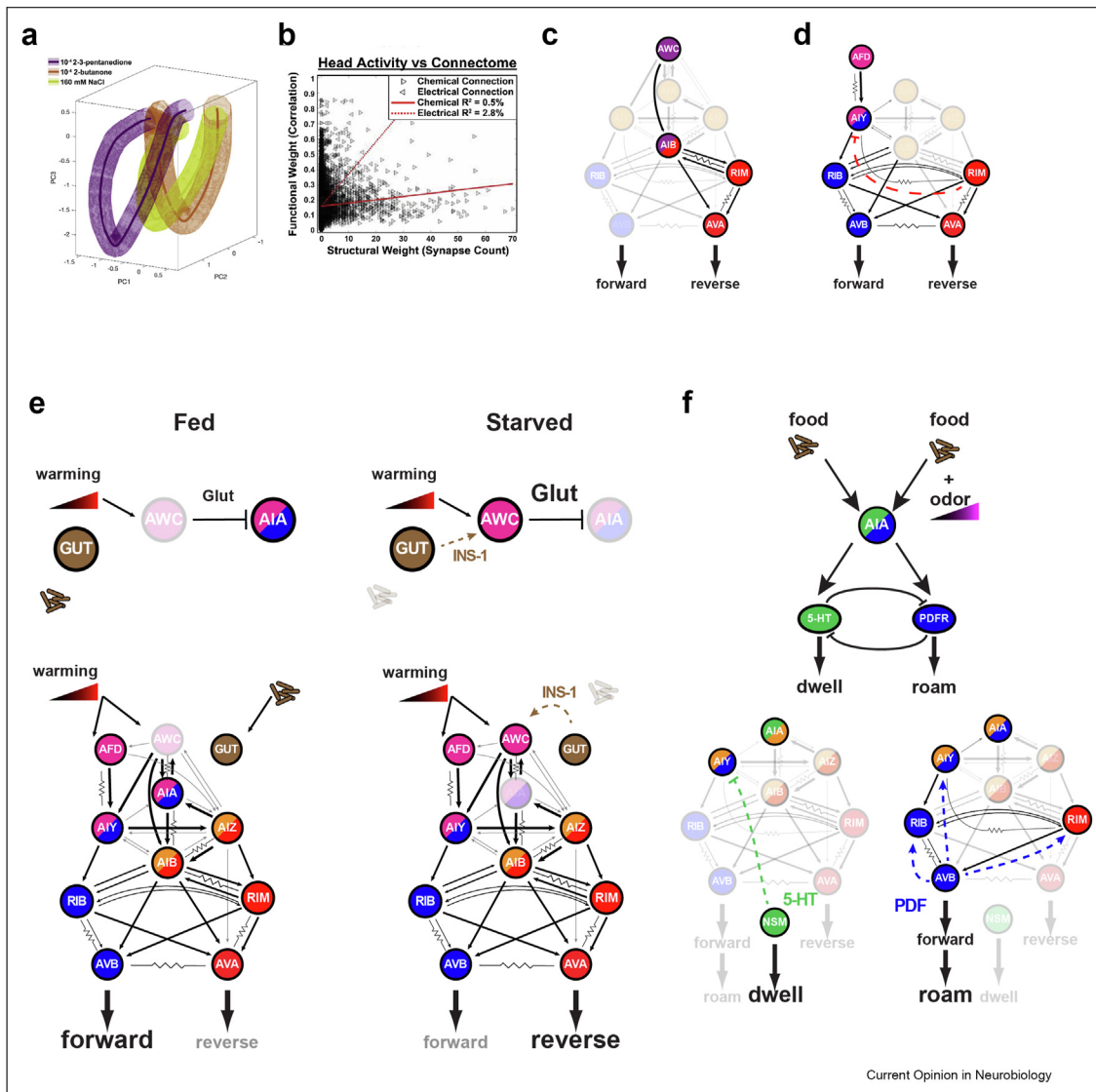
An analysis of volumetric reconstructions across distinct hermaphrodite animals found that half of the membrane contacts are largely conserved across several length scales of organization [12,13]. The brain's physical structure follows a conserved developmental program that imposes physical constraints on synaptic placement, restricting the potential connectivity of neurons to local neighborhoods. The shapes and relative

positions of neurites are largely conserved at birth, and synapse number grows in proportion to the body [14]. Most membrane contacts are stable during development, with most additional synapses strengthening existing connections, and new connections largely occurring at existing large contact areas (Figure 1a). Motor networks in particular are highly stereotyped, likely reflecting the importance of motor fidelity. However, modulatory neurons, which rely less on synaptic signaling, are the least precise in synaptic loci and number. Like the stereotypy of cellular identity, the physical geometry and contacts of neurons across the nervous system are largely conserved, with most individual variability occurring at sparse synapse loci.

Two important features of brain organization are absent from physical connectivity maps. First, the signs and strengths of the synaptic connections cannot be inferred from EM images. Second, the physical network provides no information about extra-synaptic communication. Neuromodulatory networks can be built by pairing known cellular expression of neuromodulators with that of their target receptors [15]. While the identities of neuromodulatory neurons and their targets are known in some cases, the receptors for most neuropeptides remain unknown and the list of *C. elegans* neuromodulators keeps expanding, including the recent addition of the IL-17 cytokine [16]. Of the neuromodulatory networks that have been mapped, these networks do not overlap well with the chemical and electrical synaptic networks. For example, most neurons that express serotonin receptors do not share synapses with serotonin-producing neurons. “Hub” neurons in the chemical and synaptic networks are not necessarily hub neurons in neuromodulatory networks. However, a small set of neurons like RIM appear to be “hub” neurons in all three networks [15].

It is not possible to determine the flow of information through a nervous system based solely on physical synaptic connectivity. However, large-scale calcium imaging approaches are beginning to reveal the flow of information in the *C. elegans* network. These studies have utilized a nuclear-localized GCaMP that allows densely-packed neurons to be easily segmented. These recordings cannot recover local calcium signals in neurites [17] and lack the temporal resolution of voltage recordings, but have still provided important insights into *C. elegans* brain-wide dynamics. Brain-wide imaging of most anterior neurons in restrained worms has revealed a high degree of spontaneous correlated activity across the brain that is altered in response to sensory stimuli [18]. Based on comparisons to neural activity in moving animals, most of this ongoing activity seems to correlate with motor patterns of the worm, which can largely be represented as a two-lobed manifold by three principal components of neuron population activities (Figure 1b–d) [19]. Different regions of this manifold represent different action sequences. Indeed, brain-wide imaging in freely-moving worms has

Figure 2



Sensory and Neuromodulatory Context Influence Interneuron Correlations with Motor Program. **a**. The path global neuronal dynamics take through PCA-reduced space is strongly influenced by sensory context. Each colored volume represents the average phase trajectory of whole-brain neuronal activity during sensory stimulation with 2–3 pentanedione (purple), 2-butanone (orange), or NaCl (yellow). Image is reproduced from Yemini et al. [23] **b**. Neuronal activity correlates poorly with chemical and electrical synapse connectivity. Image is reproduced from Yemini et al. [23] **c**. The AIB interneuron exhibits activity that correlates with sensory-induced activity from AWC, and/or reverse motor activity mediated by synaptic input from RIM. **d**. The AIY interneuron exhibits activity that correlates with sensory-induced activity from AFD, and/or forward motor activity mediated by neuromodulatory inhibition from RIM. **e**. AIA activity is strongly influenced by both sensory and food context. Under well-fed conditions, AWC activity does not correlate with warming gradients, and in turn does not inhibit AIA activity via glutamate. Under starved conditions, AWC is sensitized to warm gradients via INS-1 signaling from the gut. Increased AWC activity silences AIA, which in turn decreases forward drive. **f**. AIA drives dwelling in the presence of food and the absence of odor gradients. Its activity is tightly coupled to the serotonergic neuron NSM which inhibits AIY and other MOD-1 expressing neurons. However, in the presence of odor gradients while on food, AIA drives roaming behavior. In this context, AIA activity is tightly coupled to forward interneuron activity.

Interneuron coupling is influenced by neuromodulatory signaling

Interneurons in particular appear to be more plastic in their functional coupling with other neurons. While sensory neurons have fairly stereotyped responses to sensory cues, and motor neurons reliably produce

behavioral responses, interneurons integrate information from a diversity of presynaptic partners with varying degrees of correlation. AIB is an interneuron important for relaying sensory information from the olfactory neuron AWC to the motor circuit (Figure 2c). On average, the activity of AIB correlates with sensory

input, but most of its activity is driven by the motor circuit through its synapses with the interneuron RIM. In the absence of RIM activity, AIB activity is more strongly correlated with sensory input and, in turn, causes the motor circuit to be more strongly coupled to AWC [25]. The role of RIM in maintaining motor states that influence sensory perception also occurs in AIY, another interneuron which encodes both thermosensory information from AFD and the motor state of the animal (Figure 2d). Here too, RIM maintains the motor drive by providing motor feedback to AIY, which in turn influences sensory perception. This feedback maintains the motor state of the animal, and makes it less susceptible to transient temperature fluctuations [26]. However, unlike AIB, RIM inhibits AIY through neuromodulatory signaling. In addition to being a hub of the physical synaptic network, RIM is also a hub of the neuromodulatory network [15], and likely plays a dual role in coordinating the activity of numerous neurons synaptically (like AIB) and extra-synaptically (like AIY). Thus, RIM may play a pivotal role in coordinating dynamics across the network of forward- and reverse-active neurons.

A more explicit role of the influence of neuromodulatory state on interneuron coupling can be observed with the interneuron AIA. Most of its presynaptic partners are sensory neurons, and its activity is strongly driven by coincident activities of these neurons [27]. However, the functional coupling of AIA to these neurons is highly dynamic. Feeding state does not alter the AFD-AIY thermotaxis circuit, but it does alter temperature-mediated behavior (Figure 2e) [28]. Under fed conditions, AIA has tonic activity that is permissive for AFD-AIY driven thermotaxis. Even though AFD is the primary thermosensor, other chemosensory neurons like AWC can become temperature-responsive under starved conditions due to INS-1 insulin-like peptide signaling from the gut. The increase in temperature-mediated AWC activity increases glutamate-mediated inhibition of AIA, which in turn influences behavior by causing a decrease in forward runs. Thus, non-neuronal tissues like the gut can strongly influence neuronal couplings, and in turn, behavior. This likely evolved to alter the relative saliency of sensory cues based on food availability, a critical contextual cue for the animal [29]. Under starved conditions, chemotaxis toward food takes precedence over thermotaxis toward optimal temperatures. Given the widespread neuronal expression of the DAF-2 insulin receptor, it is possible that there may be similar state-dependent changes in other components of the sensory circuits.

In addition to neuromodulatory signaling from the gut to the brain that influences behavior, olfactory perception by the brain also regulates gut metabolism. Odor detection by sensory neurons leads to decreased octopamine release by the neuron RIC, which in turn leads to a

decrease in AMPK activity in the gut [30]. Octopamine is the invertebrate analog of norepinephrine, and AMPK is a key metabolic regulator which activates glycolysis and fatty-acid metabolism. This study and others [31,32] highlight how communication along the gut–brain axis is bidirectional: neuromodulatory signaling from the gut influences behavior, but neuromodulators released by the brain influence gut metabolism.

The dual roles of chemical synapses and neuromodulation on functional connectivity can also be seen in the timescales of neuronal coupling. When animals are actively moving in the absence of food, AIA has transient activity that is positively correlated with the forward state, which is consistent with its role in driving forward behavior in response to attractive sensory input. However, during dwelling states on food, AIA activity is more sustained and correlated with the dwelling-promoting serotonergic neuron NSM (Figure 2f) [33]. Alternatively, odor gradients in the presence of food can trigger AIA to promote roaming, and return to being highly correlated with the forward motor circuit. The ability of AIA to exert opposing effects on behavior is enabled by its dual excitatory outputs to serotonin and PDF neurons that mutually inhibit one another to promote dwelling and roaming states, respectively (see below for further details). Thus, even though AIA is activated by food odors in both states, its influence on speed (fast timescale) is influenced by the roaming/dwelling state of the animal (slow timescale). This change in neuronal coupling reflects the changing priorities of the animals in different behavioral states: food odors during roaming provide a cue for navigation to a distal target, whereas food odors during dwelling and food ingestion indicate that the animal is already in a plentiful food resource. These studies reveal that despite a largely invariant physical network, the functional network is dynamic, and influenced by extra-synaptic signaling that alters how interneurons couple to both sensory and motor circuits.

The internal states of the *C. elegans* nervous system

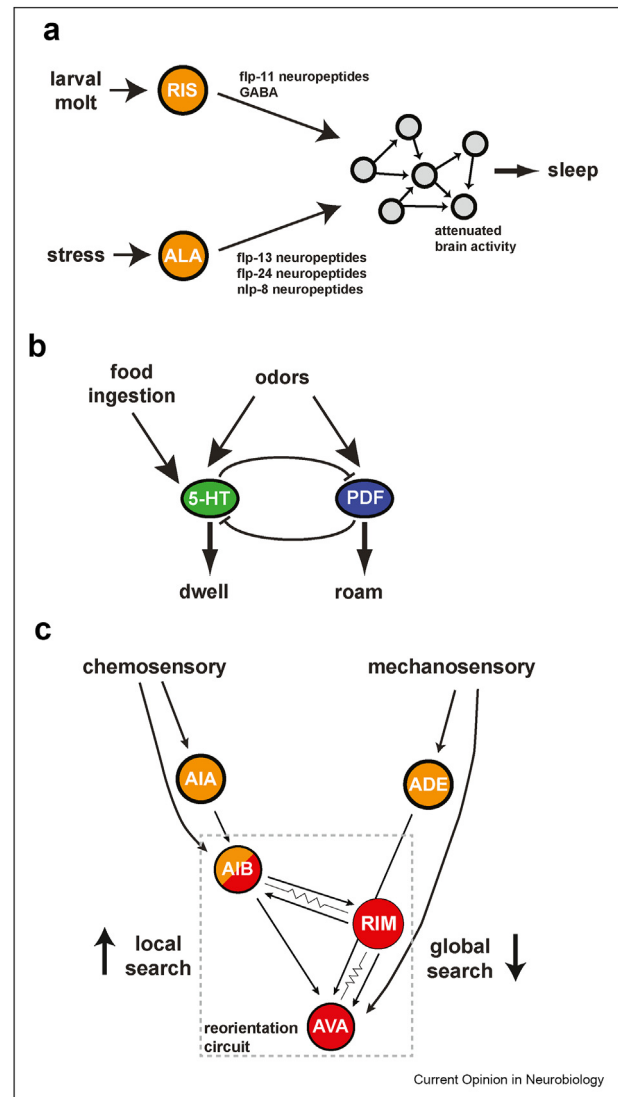
The internal state of an animal's nervous system can robustly alter sensorimotor processing and behavior. Although “internal state” is a somewhat loosely-defined term, here we define an internal state to be a persistent change in the function of the nervous system whose effects span multiple sensory modalities and/or motor systems. *C. elegans* exhibits a range of distinct internal states that influence functional circuit connectivity and sensorimotor transformations, such as sleep, hunger, and exploration [34]. We now review the neural mechanisms that generate these states.

The clearest example of an internal state that alters sensorimotor behavior in *C. elegans* is the sleep state. Similar to mammals, sleep in *C. elegans* can be defined

based on behavioral criteria: a cessation of movement and an increased arousal threshold, which is rapidly reversible and exhibits homeostatic buildup [35,36]. Unlike mammalian systems, there is not yet a widely-established neurophysiological definition of *C. elegans* sleep, though recent brain-wide recordings suggest that sleep is correlated with a broad inhibition of the brain-wide neural dynamics described above [37]. The increased arousal threshold appears to be due to both reduced responsiveness of sensory neurons and alterations in downstream sensory processing circuits [38]. *C. elegans* display sleep states primarily in two contexts (Figure 3a). First, they display developmentally-timed sleep (DTS) during each of their four larval molts [35]. Second, they display sickness-induced sleep (SIS) after exposure to robust stressors, such as potentially lethal levels of heat or tissue damage [39,40]. The neural control of sleep involves command neurons that act through neuropeptide release to modulate downstream circuits. DTS is associated with activation of the RIS neuron, whose release of FLP-11 neuropeptides induces behavioral quiescence [41]. In addition, PDF-1/2 and FLP-2 neuropeptides produced by sensory neurons modulate DTS [42]. On the other hand, SIS is controlled primarily by the ALA neuron that releases several neuropeptides [43,44]. The ALA-secreted neuropeptides have unique but overlapping functions in inhibiting specific motor programs of the worm, suggesting combinatorial control [43]. The control of *C. elegans* sleep by neuropeptides is reflective of the broadly important role that neuropeptide signaling plays in organizing *C. elegans* neural dynamics and behavior.

Awake *C. elegans* transition between a wide range of additional internal states. While foraging on a bacterial food source, animals abruptly switch between stable (5–60 min) “roaming” and “dwelling” behavioral states in which they display either active exploration or slow exploitation of their food source, respectively [45–47]. Roaming and dwelling animals also exhibit behavioral differences beyond locomotion, for example different levels of egg-laying and olfactory-driven behavioral responses [33,48]. Animals favor dwelling in environments with dense, nutritive food, whereas they favor roaming when there is less food, aversive sensory stimuli, or a food odor gradient from a better nearby food source [33,46,49]. Transitions between roaming and dwelling are controlled by a neural circuit consisting of a mutual inhibitory loop between neurons that make two opposing neuromodulators: serotonin, which promotes dwelling, and PDF neuropeptides, which promote roaming (Figure 3b) [33]. Optogenetic activation of the serotonergic or PDF systems can flip the behavioral state of the animal and loss of these neuromodulatory systems causes fragmented, short states [47]. Diverse sensory signals feed into this core mutual inhibitory loop. The ingestion of bacterial food activates the serotonergic neuron NSM, which extends a

Figure 3



Neural Circuits for Behavioral State Control in *C. elegans*. **a.** The RIS interneuron controls developmentally timed sleep, whereas the ALA interneuron controls stress-induced sleep. Each releases distinct neuropeptides that have causal roles in inhibiting the main *C. elegans* motor programs and thereby inducing sleep. **b.** Mutual inhibition between the neurons that produce the opposing 5-HT and PDF neuromodulators underlies bi-stable switching between roaming and dwelling behavioral states. Food ingestion activates the serotonergic system to increase dwelling, while both neuromodulatory systems receive feedforward inputs from the chemosensory system. **c.** Chemosensory and mechanosensory cues are detected by different sets of sensory neurons that feed into a reorientation circuit. Immediately after food removal, the sensory neurons are spontaneously active and strongly coupled to the reorientation circuit. As animal continue to explore without successfully finding food, sensory neurons display reduced activity and reduced coupling to the reorientation circuit.

sensory dendrite into the alimentary canal to directly sense bacterial food [50]. Olfactory processing neurons can couple to neurons in the roaming or dwelling circuit, depending on which state the animal is in [33]. Convergence of diverse sensory signals onto the core

mutual inhibitory circuit allows these states to be influenced by the environment. State-dependent coupling of sensory neurons to either roaming or dwelling neurons allows for state-dependent sensory processing.

In the absence of bacterial food, *C. elegans* display a different set of behavioral states: immediately after food removal, they exhibit Local Search (LS) in which they display a high incidence of turns that allows them to potentially re-encounter food. After several minutes, they transition to a Global Search (GS) in which they reduce their turning (Figure 3c) [51–53]. The LS state is triggered by food-detecting olfactory and mechano-sensory neurons that act in a redundant fashion on downstream processing circuits [54]. Onset of the GS state is associated with reduced activity in the sensory neurons and reduced coupling of the sensory neurons to downstream motor circuits.

The metabolism and satiety of *C. elegans* exerts a strong influence over its behavioral states. Starvation followed by re-feeding can trigger a quiescence state that is reminiscent of sleep [55]. Entry into this state is influenced by the level of fat stores in the animal [56], as well as neurohumoral signaling via insulin/DAF-2 signaling and TGF-beta/DAF-7 signaling. Alterations in TGF-beta/DAF-7 signaling during pathogenic infection further modulates behavior [57,58]. Roaming and dwelling states are also influenced by satiety: animals favor dwelling after periods of fasting [46]. This appears to involve detection of peripheral fat stores, as well as enhanced activation of the serotonergic neuron NSM by food ingestion in fasted animals [50,59–61]. Starvation also impacts other complex *C. elegans* behaviors. For example, the drive to cross an aversive sensory barrier to reach an appetitive food cue is increased by starvation due to inhibition of the tyraminerbic neuron RIM, which controls the gain of the response to the aversive cue [62]. Starvation can also change the valence of individual sensory cues, like carbon dioxide [63]. The profound effect of satiety levels on *C. elegans* behavior illustrates the strong influence that an animal's metabolic state can exert on neural circuit function.

Finally, *C. elegans* behavioral states are also influenced by developmental age and developmental history. The fraction of time that an animal roams and dwells changes in a characteristic manner as it passes through its four larval stages into adulthood [64]. These changes are influenced by a wide range of neuromodulatory pathways. Related to these changes, the reproductive status of the animal has a causal role in altering roaming and dwelling behaviors as adult animals gain the ability to lay eggs [65,66]. Early life events also shape adult behavioral states days later. Animals that are food-deprived as young larvae and pass through the dauer diapause stage

display more cautious foraging strategies as adults, consisting of reduced exploration [67]. These behavioral changes involve altered neural dynamics in a sensory processing circuit important for navigation, rather than changes at the sensory periphery. These studies add to an emerging view that the internal states of *C. elegans* represent highly integrative responses to sensory and physiological cues that endow the animal with flexible behavioral control over long time scales.

Concluding remarks

While the neuronal network itself is physically static, the functional connections between neurons are dynamic over time and across different sensory contexts. Evolving neuromodulatory signaling alters the functional state of the neuronal network, which is manifested as different behavioral states. The brain-wide dynamics that accompany these states are the subject of active investigation and initial studies are revealing different correlative structure and dynamics across states. Knowing these correlative dynamics will be critically helpful to parameterize computational models of network function. A variety of computational approaches exist, from biophysical models that model each neuron and synapse [68], to Bayesian models that capture the correlative relationships between neurons [69], to models that combine neurons into state spaces based on the largest principle components of neuronal dynamics [70]. All of these models provide helpful insights into network function at different resolutions of network simplification. However, these models fit their parameters based on observed dynamics and functional relationships, and these empirical measurements remain very limited in the field. Further studies of brain-wide activity states will provide useful output solutions to constrain models and should reveal which underlying parameters of network function are critical to producing observed changes in functional connectivity and brain-wide dynamics.

The diversity of possible solutions encoded within the physical synaptic network provides the worm with a multitude of behaviors to engage with a constantly evolving environment. By encoding multiple solutions in a single network, the worm is better able to adapt to a changing environment on both individual and evolutionary timescales. New behavioral responses can occur by altering the functional relationships of neurons that already exist. This strategy is ancient. The roles of neuromodulators such as serotonin and dopamine in controlling states like foraging, satiety, and sensory valence are conserved amongst invertebrates and vertebrates [71–73]. However, with the single-neuron resolution of the *C. elegans* network, it should be feasible to understand how these neuromodulatory signals explicitly alter network properties that may be difficult to define in larger systems.

In addition to neuromodulators like serotonin and dopamine, there are over 200 neuropeptides encoded in the *C. elegans* genome [74]. Future studies that further define the neuromodulatory connectome and reveal brain-wide dynamics during defined neuromodulatory states should provide new insights into the context dependence of *C. elegans* brain-wide activities. Studies of specific subsets of neurons, described above, are beginning to reveal mechanisms for context-dependent behavior, but this work should be contextualized in a broader framework of network-wide dynamics. The number of possible internal states encoded by neuromodulators, and their behavioral consequences, are likely to be vast, and a rich topic for future research.

Conflict of interest statement

Nothing declared.

Acknowledgments

We thank members of the Flavell and Gordus labs for critical comments on the manuscript. S.W.F. acknowledges funding from NIH (NS104892 and GM135413), NSF (1845663), the JPB Foundation, the Alfred P. Sloan Foundation, the Brain Research Foundation, and the McKnight Scholars Program. A.G. acknowledges funding from NIH (GM124883).

References

Papers of particular interest, published within the period of review, have been highlighted as:

* of special interest

** of outstanding interest

1. Niell CM, Stryker MP: **Modulation of visual responses by behavioral state in mouse visual cortex.** *Neuron* 2010, **65**:472–479.
2. Goard M, Dan Y: **Basal forebrain activation enhances cortical coding of natural scenes.** *Nat Neurosci* 2009, **12**:1444–1449.
3. Polack P-O, Friedman J, Golshani P: **Cellular mechanisms of brain state-dependent gain modulation in visual cortex.** *Nat Neurosci* 2013, **16**:1331–1339.
4. Brincat SL, Siegel MR, von Nicolai C, Miller EK: **Gradual progression from sensory to task-related processing in cerebral cortex.** *Proc Natl Acad Sci U S A* 2018, **115**:E7202–E7211.
5. Moore T, Zirnsak M: **Neural mechanisms of selective visual attention.** *Annu Rev Psychol* 2017, **68**:47–72.
6. White JG, Southgate E, Thomson JN, Brenner S: **The structure of the nervous system of the nematode *Caenorhabditis elegans*.** *Philos Trans R Soc Lond B Biol Sci* 1986, **314**:1–340.
7. Cook SJ, et al.: **Whole-animal connectomes of both *Caenorhabditis elegans* sexes.** *Nature* 2019, **571**:63–71.
8. Varshney LR, Chen BL, Paniagua E, Hall DH, Chklovskii DB: **Structural properties of the *Caenorhabditis elegans* neuronal network.** *PLoS Comput Biol* 2011, **7**, e1001066.
9. Towson EK, Vértes PE, Ahnert SE, Schafer WR, Bullmore ET: **The rich club of the *C. elegans* neuronal connectome.** *J Neurosci Off J Soc Neurosci* 2013, **33**:6380–6387.
10. Chalfie M, et al.: **The neural circuit for touch sensitivity in *Caenorhabditis elegans*.** *J Neurosci Off J Soc Neurosci* 1985, **5**: 956–964.
11. Roberts WM, et al.: **A stochastic neuronal model predicts random search behaviors at multiple spatial scales in *C. elegans*.** *Elife* 2016, **5**, e12572.
12. Brittin CA, Cook SJ, Hall DH, Emmons SW, Cohen N: **A multi-scale brain map derived from whole-brain volumetric reconstructions.** *Nature* 2021, **591**:105–110.
13. Moyle MW, et al.: **Structural and developmental principles of neuropil assembly in *C. elegans*.** *Nature* 2021, **591**:99–104.
14. Witvliet D, et al.: **Connectomes across development reveal principles of brain maturation.** *Nature* 2021, **596**:257–261.
15. Bentley B, et al.: **The multilayer connectome of *Caenorhabditis elegans*.** *PLoS Comput Biol* 2016, **12**, e1005283.
16. Chen C, et al.: **IL-17 is a neuromodulator of *Caenorhabditis elegans* sensory responses.** *Nature* 2017, **542**:43–48.
17. Hendricks M, Ha H, Maffey N, Zhang Y: **Compartmentalized calcium dynamics in a *C. elegans* interneuron encode head movement.** *Nature* 2012, **487**:99–103.
18. Schrödel T, Prevedel R, Aumayr K, Zimmer M, Vaziri A: **Brain-wide 3D imaging of neuronal activity in *Caenorhabditis elegans* with sculpted light.** *Nat Methods* 2013, **10**:1013–1020.
19. Kato S, et al.: **Global brain dynamics embed the motor command sequence of *Caenorhabditis elegans*.** *Cell* 2015, **163**: 656–669.
20. Nguyen JP, et al.: **Whole-brain calcium imaging with cellular resolution in freely behaving *Caenorhabditis elegans*.** *Proc Natl Acad Sci U S A* 2016, **113**:E1074–E1081.
21. Venkatachalam V, et al.: **Pan-neuronal imaging in roaming *Caenorhabditis elegans*.** *Proc Natl Acad Sci U S A* 2016, **113**: E1082–E1088.
22. Hallinen KM, et al.: **Decoding locomotion from population neural activity in moving *C. elegans*.** *Elife* 2021, **10**, e66135.
23. Yemini E, et al.: **NeuroPAL: a multicolor atlas for whole-brain neuronal identification in *C. elegans*.** *Cell* 2021, **184**:272–288.
24. Susoy V, et al.: **Natural sensory context drives diverse brain-wide activity during *C. elegans* mating.** *Cell* 2021, **184**: 5122–5137. e17.
25. Gordus A, Pokala N, Levy S, Flavell SW, Bargmann CI: **Feedback from network states generates variability in a probabilistic olfactory circuit.** *Cell* 2015, **161**:215–227.
26. Ji N, et al.: **Corollary discharge promotes a sustained motor state in a neural circuit for navigation.** *Elife* 2021, **10**, e68848.
27. Dobosiewicz M, Liu Q, Bargmann CI: **Reliability of an interneuron response depends on an integrated sensory state.** *Elife* 2019, **8**, e50566.
28. Takeishi A, Yeon J, Harris N, Yang W, Sengupta P: **Feeding state functionally reconfigures a sensory circuit to drive thermosensory behavioral plasticity.** *Elife* 2020, **9**, e61167.
29. Kim DH, Flavell SW: **Host-microbe interactions and the behavior of *Caenorhabditis elegans*.** *J Neurogenet* 2020, **34**: 500–509.
30. Zhang B, Jun H, Wu J, Liu J, Xu XZS: **Olfactory perception of food abundance regulates dietary restriction-mediated longevity via a brain-to-gut signal.** *Nat Aging* 2021, **1**:255–268.
31. Hussey R, et al.: **Pheromone-sensing neurons regulate peripheral lipid metabolism in *Caenorhabditis elegans*.** *PLoS Genet* 2017, **13**, e1006806.
32. Hussey R, et al.: **Oxygen-sensing neurons reciprocally regulate peripheral lipid metabolism via neuropeptide signaling in *Caenorhabditis elegans*.** *PLoS Genet* 2018, **14**, e1007305.

33. Ji N, *et al.*: **A neural circuit for flexible control of persistent behavioral states.** *Elife* 2021, **10**, e62889.
This study utilized large-scale calcium imaging in defined populations of neurons, together with neural circuit perturbations, to uncover the functional logic of a neural circuit controlling opposing roaming and dwelling states.
34. Flavell SW, Raizen DM, You Y-J: **Behavioral States.** *Genetics* 2020, **216**:315–332.
35. Raizen DM, *et al.*: **Lethargus is a *Caenorhabditis elegans* sleep-like state.** *Nature* 2008, **451**:569–572.
36. Van Buskirk C, Sternberg PW: **Epidermal growth factor signaling induces behavioral quiescence in *Caenorhabditis elegans*.** *Nat Neurosci* 2007, **10**:1300–1307.
37. Nichols ALA, Eichler T, Latham R, Zimmer M: **A global brain state underlies *C. elegans* sleep behavior.** *Science* 2017, **356**, eaam6851.
38. Cho JY, Sternberg PW: **Multilevel modulation of a sensory motor circuit during *C. elegans* sleep and arousal.** *Cell* 2014, **156**:249–260.
39. Hill AJ, Mansfield R, Lopez JMNG, Raizen DM, Van Buskirk C: **Cellular stress induces a protective sleep-like state in *C. elegans*.** *Curr Biol CB* 2014, **24**:2399–2405.
40. Nelson MD, *et al.*: **FMRamide-like FLP-13 neuropeptides promote quiescence following heat stress in *Caenorhabditis elegans*.** *Curr Biol CB* 2014, **24**:2406–2410.
41. Turek M, Besseling J, Spies J-P, König S, Bringmann H: **Sleep-active neuron specification and sleep induction require FLP-11 neuropeptides to systemically induce sleep.** *Elife* 2016, **5**, e12499.
42. Chen D, Taylor KP, Hall Q, Kaplan JM: **The neuropeptides FLP-2 and PDF-1 act in concert to arouse *Caenorhabditis elegans* locomotion.** *Genetics* 2016, **204**:1151–1159.
43. Nath RD, Chow ES, Wang H, Schwarz EM, Sternberg PWC: **Elegans stress-induced sleep emerges from the collective action of multiple neuropeptides.** *Curr Biol CB* 2016, **26**:2446–2455.
44. Iannaccone MJ, *et al.*: **The RFamide receptor DMSR-1 regulates stress-induced sleep in *C. elegans*.** *Elife* 2017, **6**, e19837.
45. Fujiwara M, Sengupta P, McIntire SL: **Regulation of body size and behavioral state of *C. elegans* by sensory perception and the EGL-4 cGMP-dependent protein kinase.** *Neuron* 2002, **36**:1091–1102.
46. Ben Arous J, Laffont S, Chatenay D: **Molecular and sensory basis of a food related two-state behavior in *C. elegans*.** *PLoS One* 2009, **4**, e7584.
47. Flavell SW, *et al.*: **Serotonin and the neuropeptide PDF initiate and extend opposing behavioral states in *C. elegans*.** *Cell* 2013, **154**:1023–1035.
48. Cermak N, *et al.*: **Whole-organism behavioral profiling reveals a role for dopamine in state-dependent motor program coupling in *C. elegans*.** *Elife* 2020, **9**, e57093.
49. Chew YL, *et al.*: **An afferent neuropeptide system transmits mechanosensory signals triggering sensitization and arousal in *C. elegans*.** *Neuron* 2018, **99**:1233–1246. e6.
50. Rhoades JL, *et al.*: **ASICs mediate food responses in an enteric serotonergic neuron that controls foraging behaviors.** *Cell* 2019, **176**:85–97. e14.
This study clarified how the ingestion of food activates a key serotonergic neuron, NSM, which elicits a behavioral state change to allow for exploitation of a food patch.
51. Gray JM, Hill JJ, Bargmann CI: **A circuit for navigation in *Caenorhabditis elegans*.** *Proc Natl Acad Sci U S A* 2005, **102**:3184–3191.
52. Wakabayashi T, Kitagawa I, Shingai R: **Neurons regulating the duration of forward locomotion in *Caenorhabditis elegans*.** *Neurosci Res* 2004, **50**:103–111.
53. Hills T, Brockie PJ, Maricq AV: **Dopamine and glutamate control area-restricted search behavior in *Caenorhabditis elegans*.** *J Neurosci Off J Soc Neurosci* 2004, **24**:1217–1225.
54. López-Cruz A, *et al.*: **Parallel multimodal circuits control an innate foraging behavior.** *Neuron* 2019, **102**:407–419. e8.
This study mapped out the neural circuit that allows recent food availability to alter the behavioral state of the animal from a local to global search as time off food progresses.
55. You Y, Kim J, Raizen DM, Avery L: **Insulin, cGMP, and TGF-beta signals regulate food intake and quiescence in *C. elegans*: a model for satiety.** *Cell Metabol* 2008, **7**:249–257.
56. Hyun M, *et al.*: **Fat metabolism regulates satiety behavior in *C. elegans*.** *Sci Rep* 2016, **6**:24841.
57. Meisel JD, Panda O, Mahanti P, Schroeder FC, Kim DH: **Chemosensation of bacterial secondary metabolites modulates neuroendocrine signaling and behavior of *C. elegans*.** *Cell* 2014, **159**:267–280.
58. Hilbert ZA, Kim DH: **Sexually dimorphic control of gene expression in sensory neurons regulates decision-making behavior in *C. elegans*.** *Elife* 2017, **6**, e21166.
59. Sawin ER, Ranganathan R, Horvitz HRC: **Elegans locomotory rate is modulated by the environment through a dopaminergic pathway and by experience through a serotonergic pathway.** *Neuron* 2000, **26**:619–631.
60. Juozaityte V, *et al.*: **The ETS-5 transcription factor regulates activity states in *Caenorhabditis elegans* by controlling satiety.** *Proc Natl Acad Sci U S A* 2017, **114**:E1651–E1658.
61. Churgin MA, McCloskey RJ, Peters E, Fang-Yen C: **Antagonistic serotonergic and octopaminergic neural circuits mediate food-dependent locomotory behavior in *Caenorhabditis elegans*.** *J Neurosci Off J Soc Neurosci* 2017, **37**:7811–7823.
62. Ghosh DD, *et al.*: **Neural architecture of hunger-dependent multisensory decision making in *C. elegans*.** *Neuron* 2016, **92**:1049–1062.
63. Rengarajan S, Yankura KA, Guillermin ML, Fung W, Hallem EA: **Feeding state sculpts a circuit for sensory valence in *Caenorhabditis elegans*.** *Proc Natl Acad Sci U S A* 2019, **116**:1776–1781.
64. Stern S, Kirst C, Bargmann CI: **Neuromodulatory control of long-term behavioral patterns and individuality across development.** *Cell* 2017, **171**:1649–1662. e10.
65. Aprison EZ, Ruvinsky I: **Dynamic regulation of adult-specific functions of the nervous system by signaling from the reproductive system.** *Curr Biol CB* 2019, **29**:4116–4123. e3.
66. Aprison EZ, Ruvinsky I: **Coordinated behavioral and physiological responses to a social signal are regulated by a shared neuronal circuit.** *Curr Biol CB* 2019, **29**:4108–4115. e4.
67. Pradhan S, Quilez S, Homer K, Hendricks M: **Environmental programming of adult foraging behavior in *C. elegans*.** *Curr Biol CB* 2019, **29**:2867–2879. e4.
68. Kunert J, Shlizerman E, Kutz JN: **Low-dimensional functionality of complex network dynamics: neurosensory integration in the *Caenorhabditis Elegans* connectome.** *Phys Rev E - Stat Nonlinear Soft Matter Phys* 2014, **89**, 052805.
69. Linderman S, Nichols A, Blei D, Zimmer M, Paninski L: **Hierarchical recurrent state space models reveal discrete and continuous dynamics of neural activity in *C. elegans*.** 2019:621540. <https://www.biorxiv.org/content/10.1101/621540v1>.
70. Morrison M, Fieseler C, Kutz JN: **Nonlinear control in the nematode *C. elegans*.** *Front Comput Neurosci* 2020, **14**:616639.
71. Tecott LH: **Serotonin and the orchestration of energy balance.** *Cell Metabol* 2007, **6**:352–361.
72. Scaplen KM, Kaun KR: **Reward from bugs to bipeds: a comparative approach to understanding how reward circuits function.** *J Neurogenet* 2016, **30**:133–148.
73. Marques JC, Li M, Schaak D, Robson DN, Li JM: **Internal state dynamics shape brainwide activity and foraging behaviour.** *Nature* 2020, **577**:239–243.
74. Van Bael S, *et al.*: **A *Caenorhabditis elegans* mass spectrometric resource for neuropeptidomics.** *J Am Soc Mass Spectrom* 2018, **29**:879–889.